

DECCAN PHARMA JOURNAL SERIES

ARMS Online Publications

www.deccanpharmajournals.com

(Review Article)

Received; accepted

PATHOPHYSIOLOGY AND PHARMACOLOGICAL INTERVENTION IN THE TREATMENT OF CARDIAC HYPERTROPHY

Ramandeep Chaudhary*¹, Anil peters¹, Shaveta Gangwani¹, Tanvi Bisht², Ramica Sharma¹

1. Rayat institute of pharmacy Railmajra, Distt Nawanshahr (Punjab) India

2. Kumaun University Department of Pharmacology, Bhimtal, Distt Nainital (Uttarakhand)

ABSTRACT

Keywords:

Cardiac hypertrophy;
cardiovascular disorders,
embryogenesis

For Correspondence:

Ramandeep Chaudhary

De Rayat institute of
pharmacy Railmajra, Distt
Nawanshahr (Punjab) India

E-mail:

raman8pharma@gmail.com

Cardiovascular disorders are the major cause of mortality and morbidity. One of the major disorders that are associated with mortality is cardiac hypertrophy. Cardiac hypertrophy is basically the enlargement of the heart muscles characterized by increased thickness of the heart muscle. Depending upon the various factors it may pathological and physiological. Cardiac hypertrophy develops in response to hemodynamic overload that is a major predictor for the development of coronary heart disease and heart failure. Pressure overload induces concentric hypertrophy characterized by wall thickening without significant chamber enlargement. During embryogenesis heart growth occurs primarily via proliferation of cardiac myocytes. However, soon after birth, cardiac myocytes withdraw irreversibly from the cell cycle and subsequently growth of the heart occurs mostly through hypertrophy. However, the factors that are responsible in the progression of cardiac hypertrophy are still not clear. Hence, the present review has been designed to study the various pathophysiological mechanisms and the pharmacological interventions in the management of cardiac hypertrophy.

Introduction

Cardiovascular disease is the main cause of mortality ⁽¹⁾ and the term cardiovascular disease comprises of broad spectrum of cardiac and circulatory pathologies ⁽²⁾. Cardiac hypertrophy has been defined as an increase in cardiomyocytes size that may be beneficial, adaptive (physiological) or maladaptive (pathophysiological) phenomenon to compensate the hemodynamic stress that arises due to pressure or volume overloads ⁽³⁻⁴⁾. Pressure overload results in concentric hypertrophy characterized by wall thickening without significant chamber enlargement ⁽⁴⁾.

Further, soon after birth, cardiac myocytes withdraw irreversibly from the cell cycle and subsequently growth of the heart occurs mostly through hypertrophy ⁽⁵⁾. Compensatory hypertrophy occurs in response to exercise and pregnancy ⁽⁶⁾. During pregnancy and exercise, the heart undergoes hypertrophic growth and eccentric hypertrophy to compensate the increase in cardiac output ⁽³⁾. Eccentric hypertrophy characterized by enlargement in the heart chamber with a proportional change in wall thickness ⁽⁴⁾. In high dynamic exercise, like running, eccentric hypertrophy is developed in response to high cardiac output due to volume overload whereas in static exercise, like weightlifting ⁽⁷⁻⁸⁾, chemical changes are developed that mark the initiation of hypertrophy ⁽³⁾. Pathological hypertrophy occurs in response to

abnormal stress. When maladaptive remodelling of reactive hypertrophy, persists for a long time along with ventricular dilation, it results in cardiac enlargement, wall thinning and wall stress ⁽⁶⁾. Moreover, it is followed by the loss of cardiac myocytes function ultimately leading to irreversible functional cardiac deterioration, heart failure and death. Furthermore, cardiac hypertrophy includes alterations in intercellular matrix leading to interstitial fibrosis and loss of β -adrenergic receptor responsiveness ⁽⁶⁾. Further, Phosphofructokinase and lactate dehydrogenase, key regulators of glycolytic metabolism, were also found to be upregulated in pathological hypertrophy induced by hypertension ⁽⁹⁻¹⁰⁾. It has been well reported that physiological hypertrophy is reversible and commonly occurs during maturation, pregnancy and exercise without any morbid effects on cardiac function ⁽¹¹⁻¹²⁾. Thus the present review has been designed to delineate pathophysiology and pharmacological intervention in the management of cardiac hypertrophy.

Pathophysiology and Signalling pathways

Cardiac hypertrophy is thickening of the myocardium which causes a marked reduction in size of the heart chamber, including the left and right ventricles and commonly occurs because of increased hemodynamic demand. Cardiac hypertrophy is generally aggregated by hypertension

⁽¹³⁾, heart valve stenosis ⁽¹⁴⁾ and heart failure ⁽¹³⁾. The renin angiotensin system (RAS) had been reported play a pivotal role in the pathogenesis of hypertension, a major inducer of hypertrophy shown in fig-1. ⁽¹⁵⁾. It has been well reported that heart failure too plays an important role in pathogenesis of hypertrophy ⁽¹⁶⁾. Both cyclooxygenase-2 (COX-2) and PI3K/Akt have also been associated in the progression of cardiac hypertrophy ⁽¹⁷⁾ and overexpression of COX-2 leads to cardiac deterioration ⁽¹⁸⁻¹⁹⁻²⁰⁾. Peroxisome proliferator-activated receptors (PPAR's) are also implicated in the pathogenesis of cardiac hypertrophy ⁽¹⁰⁾. PPAR α is a target of ERK1/2 which gets activated in settings of pathological cardiac hypertrophy ⁽²¹⁾. However, the role of PPAR γ in the pathogenesis of cardiac hypertrophy is not well understood ⁽²²⁾. Further, various studies indicate the pivotal role of calcineurin in the development of pressure overload-induced cardiac hypertrophy ⁽²³⁻²⁴⁾. NGF1A-binding protein (Nab1) is a transcriptional repressor which appears to be a specific regulator of pathological cardiac hypertrophy ⁽¹⁰⁻²⁵⁾.

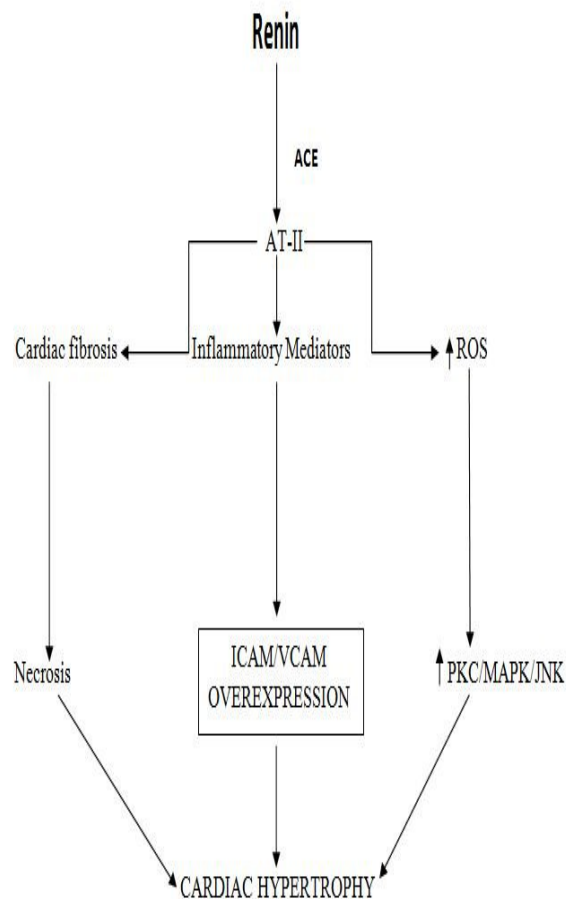


Figure 1. Role of various mediators in mediating cardiac hypertrophy where ACE = Angiotensinogen converting enzyme; ROS = Reactive oxygen species; PKC = Protein kinase C; MAPK = Mitogen activated protein kinase; JNK = Jan nuclear kinase, ICAM = Intracellular adhesion molecule, VCAM = Vascular adhesion molecule.

A recent study has indicated the role of estrogens in both physiological and pathological cardiac hypertrophy indicated in Fig-2⁽²⁶⁻²⁷⁻²⁸⁻²⁹⁾. Cardiac hypertrophy was also observed in patient with hyperthyroidism that results due to increased activity of sympathetic nervous system and increase level of RAS.⁽¹⁰⁻¹⁵⁾ Increase activity of NADPH oxidase results in cardiac hypertrophy as it is a key source for the generation of reactive oxygen species (ROS) which have been shown to play a portal role in the development of pathological hypertrophy⁽³⁰⁻³¹⁾. Further, in cardiac hypertrophy there is overexpression of C-reactive protein (C-RP)⁽³²⁾. Moreover, oxidative and nitrosative stresses plays a vital role in inducing cardiac hypertrophy and result in the generation of reactive oxygen species (ROS). ROS stimulates various signalling pathways like tyrosine kinase Src, GTP-binding protein Ras, protein kinase C, mitogen-activated protein kinase (MAPK), and Jun-nuclear kinase (JNK)⁽³³⁻³⁴⁾. Further, Tumor necrosis factor- α (TNF- α), a proinflammatory cytokine has been implicated in the pathogenesis of cardiovascular disorders⁽³⁵⁻³⁶⁻³⁷⁻³⁸⁾. However, the prolonged exposure to high levels of TNF- α is associated with cardiac dysfunction and hypertrophy of the heart⁽³⁹⁾. The TNF superfamily such as TNF-related apoptosis inducing ligand (TRAIL), osteoprotegerin (OPG), receptor activator of NF κ B ligand (RANKL), CD40L and CD27L were upregulated in heart failure patients⁽⁴⁰⁾. These mediators contributed to inflammation,

cardiac remodeling and apoptotic cell death in the failing heart⁽⁴⁰⁻⁴¹⁾. The RANKL increased the expression of matrix metalloproteinase (MMP) in cardiomyocytes and produced cardiac remodelling⁽⁴²⁾. C-reactive protein (CRP), a cytokine and a member of the class of acute-phase reactants, was significantly elevated during the inflammatory process and was associated with cardiovascular risk and the development of heart failure⁽⁴³⁻⁴⁴⁾.

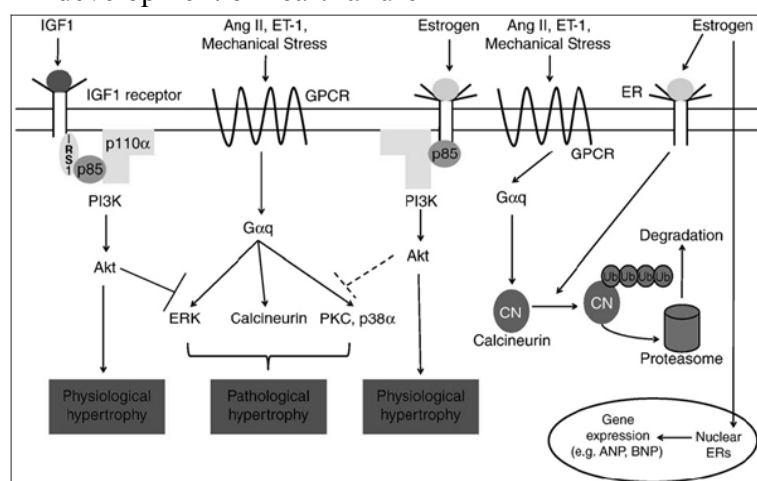


Figure 2. Signaling cascades involved in estrogen-mediated cardiac hypertrophic responses.

Ang II: angiotensin II, ANP: Atrial natriuretic peptide, BNP: B-type natriuretic peptide, CN: calcineurin, ER: estrogen receptor, ERK: extracellular regulated kinase, ET-1: endothelin-1, GPCR: G protein-coupled receptor, IGF1: insulin-like growth factor 1, IRS1: insulin receptor substrate 1, PI3K: phosphoinositide 3-kinase, PKC: protein kinase C, Ub: ubiquitin.

Pharmacological interventions (Table 1)

Anti-TNF- α therapy

Pro-inflammatory cytokines has been reported to play a vital role in the development of cardiac hypertrophy and remodelling. In patients with CHF, there is up regulation expression of pro-inflammatory cytokines, including tumour necrosis factor α (TNF- α), interleukins (IL) ⁽⁴⁵⁾. Various studies indicated the role of IL-6 in the progression of cardiovascular disease. So, treatment with anti TNF- α attenuates cardiac hypertrophy. Etanercept have been reported to inhibit the TNF- α ⁽⁴⁶⁾. Chronic treatments with a low dose of the cholinesterase inhibitor neostigmine or the muscarinic agonist pilocarpine attenuated cardiac hypertrophy and ventricular dysfunction induced by pressure overload. These treatments attenuated the increased TNF- α expression and simultaneously increased IL-10 expression in the hearts with cardiac pressure overload. Moreover, the cholinergic agonist pilocarpine elicits a direct inhibition on increased TNF- α production and hypertrophic responses in primary cultured cardiac cells. Pharmacologically, neostigmine inhibits acetylcholine inactivation and thus prolongs the excitation of cholinergic synapses. Pilocarpine stimulates postsynaptic muscarinic cholinergic receptors. Both agents have been reported to reduce the levels of the pro-inflammatory cytokine TNF- α , increased the levels of the anti-inflammatory cytokine IL-10 and attenuated cardiac hypertrophy ⁽⁴⁷⁾.

Poly (ADP-ribose) polymerase

Poly (ADP-ribose) polymerase (PARP) is a DNA repairing enzyme present in nuclei and mitochondria of various cells including cardiac myocytes. PARP activation upregulated various transcription factors and hypertrophic genes ⁽⁴⁸⁻⁴⁹⁾, which has been reported to produce pathological cardiac hypertrophy ⁽⁵⁰⁾. The PARP-mediated induction of inflammatory cytokines and apoptotic cell death play a major role in the progression of cardiac hypertrophy and decompensated heart failure ⁽⁵⁰⁻⁵¹⁾. Inhibition of PARP by 3-aminobenzamide and 5-aminoisoquinolinone markedly prevented pressure over load-induced cardiac dysfunction and pathological cardiac hypertrophy ⁽⁵²⁻⁵³⁾. Thus PARP over-activation plays a pivotal role in the transformation of pathological cardiac hypertrophy into heart failure ⁽⁵⁴⁾.

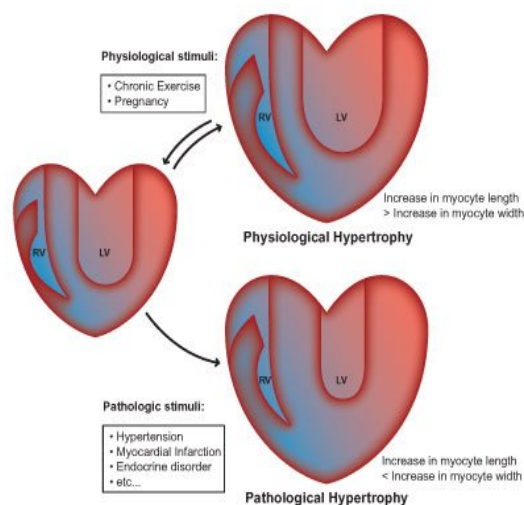


Figure 3. Pathological and physiological hypertrophic responses to stimuli.

Glucagon-like peptide-1

Both pre-clinical and clinical studies indicate that proglucagon-derived peptide, Glucagon-like peptide-1 (GLP-1), exerts favourable actions on cardiovascular function⁽⁵⁵⁾. GLP-1 receptor (GLP-1R) is expressed in the heart⁽⁵⁶⁾ and GLP-1R agonists directly activate cardiomyocyte signaling pathways⁽⁵⁷⁾. Moreover, it has been well documented that Glucagon-like peptide-1 receptor (GLP-1R) agonists are used to treat type-2 diabetes and transient GLP-1 administration improved cardiac function in humans following acute myocardial infarction (MI) and percutaneous revascularization⁽⁵⁸⁾. Liraglutide increased cyclic AMP formation and reduced the extent of caspase-3 activation in cardiomyocytes in a GLP-1R-dependent. Various studies indicated the Liraglutide drug administration induced changes in the expression of cardioprotective proteins in the normal non-atherosclerotic murine heart characterized by phosphorylation of Akt and GSK3 β and increased expression of Nrf2, PPAR- β/δ and HO-1⁽⁵⁸⁾.

Antioxidant

EPIGALLOCATECHIN-3-GALLATE

Epigallocatechin-3 gallate (EGCG) is the major catechin derived from green tea, has been found to have protective effects on the cardiovascular system⁽⁵⁹⁾. It has been reported to have anti-inflammatory effects⁽⁶⁰⁾, lower serum cholesterol levels and thus implicated in the management of reducing atherosclerosis⁽⁶¹⁾. EGCG was

found to have antioxidant properties⁽⁶²⁾ and is thus implicated in the management of oxidative stress-related diseases such as ischemic heart diseases (IHD). Further, cardiac hypertrophy results in the increase in blood pressure and wall stress which is regulated by a number of intracellular signal transduction pathways including mitogen-activated protein kinase (MAPK), Janus kinase/signal transducers, activators of transcription (JAK/STAT), calcineurin, serine/threonine kinase, Akt and its downstream target, glycogen synthase kinase-3 β ⁽⁶³⁾. The platelet derived growth factor (PDGF)-BB which is inhibited by EGCG induce intracellular Ca²⁺ increase and extracellular signal regulated kinase (ERK) activation in vascular smooth muscle cells (VSMCs)⁽⁶⁴⁾ and that EGCG could prevent Ang II-induced VSMCs hypertrophy through blocking c-Jun N-terminal kinases (JNKs)⁽⁶⁵⁾. Moreover, EGCG inhibit the PI3K/Akt and Erk1/2 pathways. Therefore it seems that EGCG can inhibit the signal pathways that regulate cardiac hypertrophy.

Tetrandrine

Tetrandrine is a bisbenzylisoquinoline alkaloid isolated from the Chinese medicinal herb-root of *Stephania tetrandra* S Moore, which was traditionally used as an anti-inflammatory, antipyretic and analgesic herb⁽⁶⁶⁻⁶⁷⁾. Tetrandrine, a naturally occurring calcium antagonist having properties like anti-inflammatory, antioxidant and anti-fibrogenetic, has

been used clinically in the management of cardiovascular diseases such as

hypertension and arrhythmia and cardiac hypertrophy⁽⁶⁸⁾.

Table 1. Novel pharmacological interventions that prevent the progression of pathological cardiac hypertrophy and heart failure.

Sr.no	Drug	Pharmacological class	Mechanism of drug in pathological cardiac hypertrophy
1	Etanercept	TNF- α -inhibitor	TNF- α -inhibition
2	Neostigmine, Pilocarpine	Cholinomimetic	Downregulation of TNF- α expression
3	3-aminobenzamide	PARP inhibitor	PARP-inhibition
4	Liraglutide	Glucagon-like peptide	Reduction in caspase-3 activation
5	Epigallocatechin-3- gallate	Antioxidant	Inhibition of MAPK
6	Tetrandrine	Antioxidant	Inhibition of cardiac fibrosis and ERK $\frac{1}{2}$ signaling pathway

Conclusion

Cardiac hypertrophy has been regarded as a major risk factor. But no reasonable pharmacological interventions have been developed for the treatment of cardiac hypertrophy. Hence, better understanding of the signalling pathways can open the new vista for the development of new drugs.

Acknowledgement

I am highly thankful to management of Rayat Educational Trust and our director Dr. AC Rana for their cooperation and innovative ideas.

REFERENCES

1. Li HH, Willis MS, Lockyer P, et al. Atrogin-1 inhibits Akt-dependent cardiac hypertrophy in mice via ubiquitin-dependent coactivation of Forkhead proteins. *Clin Invest*. 2007; 117:3211–23.
2. Krystof V, Chamrad I, Jorda R, et al. Pharmacological Targeting of CDK9 in Cardiac Hypertrophy. *Medicinal Research Reviews*. 2010; 30:646-66.
3. Eghbali M, Deva R, Alioua A, et al. Molecular and functional signature of heart hypertrophy during pregnancy. *Circ Res*. 2005; 96:1208-1216.
4. Dorn GW, Robbins J, Sugden PH. Phenotyping hypertrophy: eschew obfuscation. *Circ Res*. 2003; 92:1171–1175.
5. Heineke J, Molkentin JD. Regulation of cardiac hypertrophy by intracellular signalling pathways. *Nat Rev Mol Cell Biol*. 2006; 7:589-600.
6. Diwan A, Gerald WD. Decompensation of Cardiac Hypertrophy: Cellular Mechanisms and Novel Therapeutic Targets. *Physiology*. 2007; 22:56–64.
7. Pluim BM, Zwinderman AH, van der Laarse A, et al. The athlete's heart. A meta-analysis of cardiac structure and function. *Circulation*. 2000; 101:336 – 344.
8. Pluim BM, Lamb HJ, Kayser HW, et al. Functional and metabolic evaluation of the athlete's heart by magnetic resonance imaging and dobutamine stress magnetic resonance spectroscopy. *Circulation*. 1998; 97:666–672.
9. Iemitsu M, Miyauchi T, Maeda S, et al. Cardiac hypertrophy by hypertension and exercise training exhibits different gene expression of enzymes in energy metabolism. *Hypertens Res*. 2003; 26:829–837.
10. Bernardo BC, Weeks KL, Pretorius L, et al. Molecular distinction between physiological and pathological cardiac hypertrophy: Experimental findings and therapeutic strategies. *Pharmacology Therapeutics*. 2010; 1-37.
11. Daniels SR, Meyer RA, Liang YC, et al. Echocardiographically determined left ventricular mass index in normal children, adolescents and young adults. *J Am Coll Cardiol*. 1988; 12:703-708.
12. Schannwell CM, Zimmermann T, Schneppenheim M, et al. Left ventricular hypertrophy and diastolic dysfunction in healthy pregnant women. *Cardiology*. 2002; 97:73-78.

13. Komuro I, Yazaki Y. Control of cardiac gene expression by mechanical stress. *Annu Rev Physiol.* 1993; 55:55-75.
14. Yu W, Chen C, Fu Y, et al. Insulin Signaling: A Possible Pathogenesis of Cardiac Hypertrophy. *Cardiovascular Therapeutics.* 2010; 28:101-5.
15. Kobori H, Ichihara A, Suzuki H, et al. Role of the renin-angiotensin system in cardiac hypertrophy-induced in rats by hyperthyroidism. *Am J Physiol.* 1997; 273:593-99.
16. Levy D, Garrison RJ, Savage DD, et al. Prognostic implications of echocardiographically determined left ventricular mass in the Framingham heart study. *N Engl J Med.* 1990; 322:1561-66.
17. Wohlschlaeger J, Schmitz KJ, Palatty J, et al. Roles of cyclooxygenase-2 and phosphorylated Akt (Thr308) in cardiac hypertrophy regression mediated by left-ventricular unloading. *J Thorac Cardiovasc Surg.* 2007; 133:37-43.
18. LaPointe MC, Mendez M, Leung A, et al. Inhibition of cyclooxygenase-2 improves cardiac function after myocardial infarction in the mouse. *Am J Physiol Heart Circ Physiol.* 2004; 286:1416-24.
19. Leng J, Han C, Demetris AJ, et al. Cyclooxygenase-2 promotes hepatocellular carcinoma cell growth through Akt activation: evidence for Akt inhibition in celecoxib-induced apoptosis. *Hepatology.* 2003; 38:756-8.
20. Catalucci D, Condorelli G. Effects of Akt on Cardiac Myocytes: Location Counts. *Circ Res.* 2006; 99:339-41.
21. Barger PM, Kelly DP. PPAR signaling in the control of cardiac energy metabolism. *Trends Cardiovasc Med.* 2000; 10:238-45.
22. Ding G, Fu M, Qin Q, et al. Cardiac peroxisome proliferator-activated receptor gamma is essential in protecting cardiomyocytes from oxidative damage. *Cardiovasc Res.* 2007; 76:269-79.
23. Lu YM, Shioda N, Han F, et al. DY-9760e inhibits endothelin-1-induced cardiomyocyte hypertrophy through inhibition of CaMKII and ERK activities. *Cardiovascular Therapeutics.* 2009; 27:17-27.
24. Chan AYM, Dolinsky VW, Soltys CLM, et al. Resveratrol Inhibits Cardiac Hypertrophy via AMP-activated Protein Kinase and Akt; *Journal of Bio Chem.* 2008; 283:24194-201.
25. Buitrago M, Lorenz K, Maass AH, et al. The transcriptional repressor Nab1 is a specific regulator of pathological Cardiac hypertrophy. *Nat Med.* 2005; 11:837-44.
26. Simoncini T, Hafezi-Moghadam A, Brazil DP, et al. Interaction of oestrogen receptor with the regulatory subunit of phosphatidylinositol-3-OH kinase. *Nature.* 2000; 407: 538-41.
27. Konhilas JP, Maass AH, Luckey SW, et al. Sex modifies exercise and cardiac adaptation in mice. *Am J Physiol Heart Circ Physiol.* 2004; 287:2768-76.
28. Du XJ, Fang L, Kiriazis H. Sex dimorphism in cardiac pathophysiology: experimental findings, hormonal mechanisms, and molecular mechanisms. *Pharmacol Ther.* 2006; 111:434-75.

29. Donaldson C, Eder S, Baker C, et al. Estrogen attenuates left ventricular and cardiomyocyte hypertrophy by an estrogen receptor-dependent pathway that increases calcineurin degradation. *Circ Res.* 2009; 104: 265-75.
30. Cave A, Grieve D, Johar S, et al. NADPH oxidase-derived reactive oxygen species in cardiac pathophysiology. *Phil Trans R Soc B.* 2005; 360:2327-34.
31. Takimoto E, Kass DA. Role of Oxidative Stress in Cardiac Hypertrophy and Remodelling. *Hypertension.* 2007; 49:241-48.
32. Wanner C. C-reactive protein Risk Prediction: Adding Cardiac Hypertrophy to the List. *American Journal of Kidney Diseases.* 2002; 40:1340-41.
33. Wei S, Rothstein EC, Fliegel L, et al. Differential MAP kinase activation and Na (+)/H (+) exchanger phosphorylation by H₂O₂ in rat cardiac myocytes. *Am J Physiol Cell Physiol* 2001; 281:1542-50.
34. Aikawa R, Nagai T, Tanaka M, et al. Reactive oxygen species in mechanical stress-induced cardiac hypertrophy. *Biochem Biophys Res Commun.* 2001; 289:901-7.
35. Matsumori A, Yamada T, Suzuki H, et al. Increased circulating cytokines in patients with myocarditis and cardiomyopathy. *Br Heart J.* 1994; 72:561-6.
36. Zhang M, Xu YJ, Saini HK, et al. TNF- α as a potential mediator of cardiac dysfunction due to intracellular Ca²⁺-overload. *Biochem Biophys Res Commun.* 2005; 327:57-63.
37. Stetson SJ, Perez-Verdia A, Mazur W, et al. Cardiac hypertrophy after transplantation is associated with persistent expression of tumor necrosis factor- α . *Circulation.* 2001; 104:676-81.
38. Zhang M, Xu YJ, Saini HK, et al. Pentoxifylline attenuates cardiac dysfunction and reduces TNF- level in ischemic-reperfused heart. *Am J Physiol Heart Circ Physiol.* 2005; 289:832-9.
39. Sarzi-Puttini P, Atzeni F, Doria A, et al. Tumor necrosis factor- α , biologic agents and cardiovascular risk. *Lupus.* 2005; 14:780-4.
40. Yndestad A. Increased gene expression of tumor necrosis factor superfamily ligands in peripheral blood mononuclear cells during chronic heart failure. *Cardiovasc Res.* 2002; 54:175-82.
41. Ueland T, Yndestad A, Oie E, et al. Dysregulated osteoprotegerin/RANK-ligand/RANK axis in clinical and experimental heart failure. *Circulation.* 2005; 111:2461-8.
42. Walsh MC, Choi Y. Biology of the TRANCE axis. *Cytokine Growth Factor Rev.* 2003; 14:251-63.
43. Bogaty P, Brophy JM, Boyer L, et al. Fluctuating inflammatory markers in patients with stable ischemic heart disease. *Arch Intern Med.* 2005; 165:221-6.
44. Satoh M, Nakamura M, Akatsu T, et al. C-reactive protein co-expresses with tumor necrosis factor-in the myocardium in human dilated cardiomyopathy. *Eur J Heart Fail.* 2005; 7:748-54.
45. Mann DL. Inflammatory mediators and the failing heart: past, present, and the

- foreseeable future. *Circ Res.* 2002; 91:988-98.
46. Horiuchi T, Mitoma H, Harashima SI, et al. Transmembrane TNF- α : structure, function and interaction with anti-TNF agents. *Rheumatology advance access.* 2010; 1-14.
 47. Freeling J, Wattier K, LaCroix C, et al. Neostigmine and pilocarpine attenuated tumour necrosis factor α expression and cardiac hypertrophy in the heart with pressure overload. *Exp Physiol.* 2007; 93.1:75-82.
 48. Oei SL, Griesenbeck J, Schweiger M, et al. Interaction of the transcription factor YY1 with human poly (ADP-ribose) transferase. *Biochem Biophys Res Commun.* 1997; 240:108-11.
 49. Kraus WL, Lis JT. PARP goes transcription. *Cell.* 2003; 113:677-83.
 50. Pillai JB, Russell HM, Raman JS, et al. Increased expression of poly (ADP-ribose) polymerase-1 contributes to caspase-independent myocyte cell death during heart failure. *Am J Physiol Heart Circ Physiol.* 2005; 288:H486-96.
 51. Virag L, Szabo C. The therapeutic potential of poly (ADP-ribose) polymerase inhibitors. *Pharmacol Rev.* 2002; 54:375-429.
 52. Balakumar P, Singh M. Possible role of poly (ADP-ribose) polymerase in pathological and physiological cardiac hypertrophy. *Methods Find Exp Clin Pharmacol.* 2006; 28:683-9.
 53. Balakumar P, Singh M. Effect of 3-aminobenzamide, an inhibitor of poly (ADP-ribose) polymerase in experimental cardiac hypertrophy. *Int J Pharmacol.* 2006; 2:543-8.
 54. Balakumar P, Jagadeesh G. Multifarious molecular signaling cascades of cardiac hypertrophy: can the muddy waters be cleared? *Pharmacological Research.* 2010; 1-19.
 55. Drucker DJ, Nauck MA. The incretin system: glucagon-like peptide-1 receptor agonists and dipeptidyl peptidase-4 inhibitors in type 2 diabetes. *Lancet.* 2006; 368:1696-1705.
 56. Ban K, Noyan-Ashraf MH, Hofer J, et al. Cardioprotective and vasodilatory actions of glucagon-like peptide 1 receptor are mediated through both glucagon-like peptides 1 receptor-dependent and -independent pathway. *Circulation.* 2008; 117:2340-50.
 57. Vila Petroff MG, Egan JM, Wang X, et al. Glucagon-like peptide-1 increases cAMP but fails to augment contraction in adult rat cardiac myocytes. *Circ Res.* 2001; 89:445-52.
 58. Noyan-Ashraf MH, Momen MA, Ban K, et al. The GLP-1R Agonist Liraglutide Activates Cytoprotective Pathways and Improves Outcomes Following Experimental Myocardial Infarction in Mice. *American Diabetes Association.* 2009; 1-22.
 59. Hao J, Kim CH, Ha TS, et al. Epigallocatechin-3 gallate prevents cardiac hypertrophy induced by pressure overload in rats. *J Vet Sci.* 2007; 8(2):121-29.
 60. Jeong WS, Kim IW, Hu R, et al. Modulatory properties of various natural chemo preventive agents on the activation of NF- κ B signaling pathway. *Pharm Res.* 2004; 21:661-670.

61. Chyu KY, Babbidge SM, Zhao X, et al. Differential effects of green tea-derived catechin on developing versus established atherosclerosis in apolipoprotein E-null mice. *Circulation*. 2004; 109:2448-53.
62. Nagai K, Jiang MH, Hada J, et al. (-)-Epigallocatechin gallate protects against NO stress-induced neuronal damage after ischemia by acting as an anti-oxidant. *Brain Res*. 2002; 956:319-22.
63. Ruwhof C, van der Laarse A. Mechanical stress-induced cardiac hypertrophy: mechanisms and signal transduction pathways. *Cardiovasc Res*. 2000; 47:23-37.
64. Ahn HY, Hadizadeh KR, Seul C, et al. Epigallocatechin-3 gallate selectively inhibits the PDGF-BB-induced intracellular signaling transduction pathway in vascular smooth muscle cells and inhibits transformation of sis-transfected NIH 3T3 fibroblasts and human glioblastoma cells (A172). *Mol Biol Cell*. 1999; 10:1093-1104.
65. Zou Y, Komuro I, Yamazaki T, et al. Cell type-specific angiotensin II-evoked signal transduction pathways: critical roles of G β γ subunit, Src family, and Ras in cardiac fibroblasts. *Circ Res*. 1998; 82:337-345.
66. Kwan CY, Achike FI. Tetrandrine and related bisbenzylisoquinoline alkaloids from medicinal herbs: cardiovascular effects and mechanisms of action. *Acta Pharmacol Sin*. 2002; 23:1057-68.
67. Yao WX, Jiang MX. Effects of tetrandrine on cardiovascular electrophysiologic properties. *Acta Pharmacol Sin*. 2002; 23:1069-74.
68. Shen DF, Tang QZ, Yan L, et al. Tetrandrine blocks cardiac hypertrophy by disrupting reactive oxygen species-dependent ERK1/2 signalling. *British Journal of Pharmacology*. 2010; 159:970-81.